



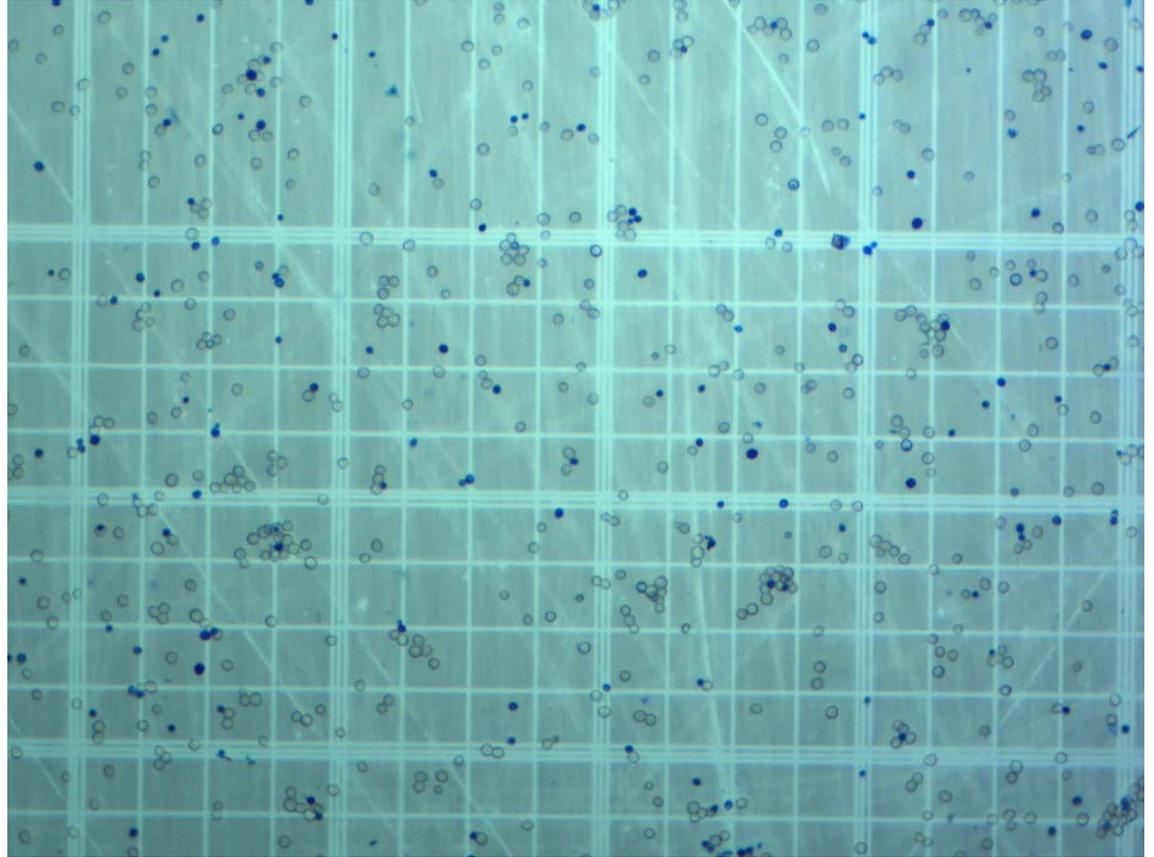
Practical Experiences with an Automated Cell Counter Using Methylene Blue in Breweries



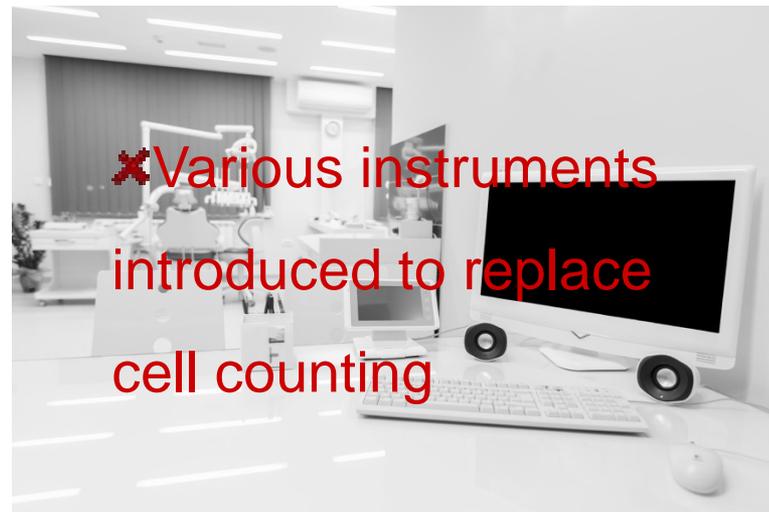
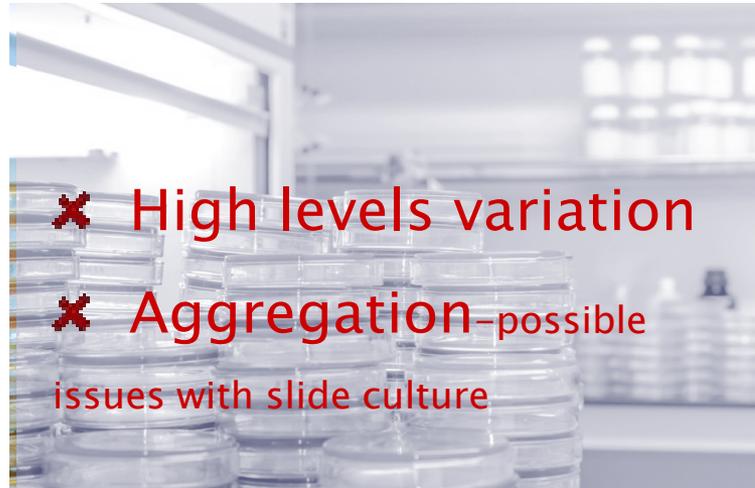
K. Thomson, J. Carvell and A. Bhat

Cell Counting

- ▶ Critical step
- ▶ Number
- ▶ Viability



Current Methods



Common and Rapid Methods for Yeast Counting and Viability Estimations

- ▶ Coulter Counter (only provides a count and not viability).
- ▶ Dielectric Spectroscopy (provides viable cell count only).
- ▶ Image Analysis with a cartridge and embedded Propidium Iodide and fluorescence microscope.
- ▶ Image analysis with addition of fluorescent stain(s) to 1 or 2 chamber slide with single or dual channels.
- ▶ Flow Cytometry.
- ▶ Image analysis with brightfield microscope/5 chamber slide and methylene blue.

Considerations by Brewery in choosing new instrument or technology

- ▶ Ease of use and level of expertise in laboratory (e.g. Process operator up to skilled microbiologist).
- ▶ Additional factors that might influence reading (e.g. Manual focussing; Extra processing of sample).
- ▶ Safety and stability of reagents and disposal.
- ▶ Cost of instrument and reagents.
- ▶ Comparability with traditional or reference data.
- ▶ Calibration and maintenance.

Purpose of the work

- ▶ Manual counts with Methylene Blue **Gold Standard**
- ▶ Inter-operator.
- ▶ Time consuming.
- ▶ Aber Countstar has been recently released into market.

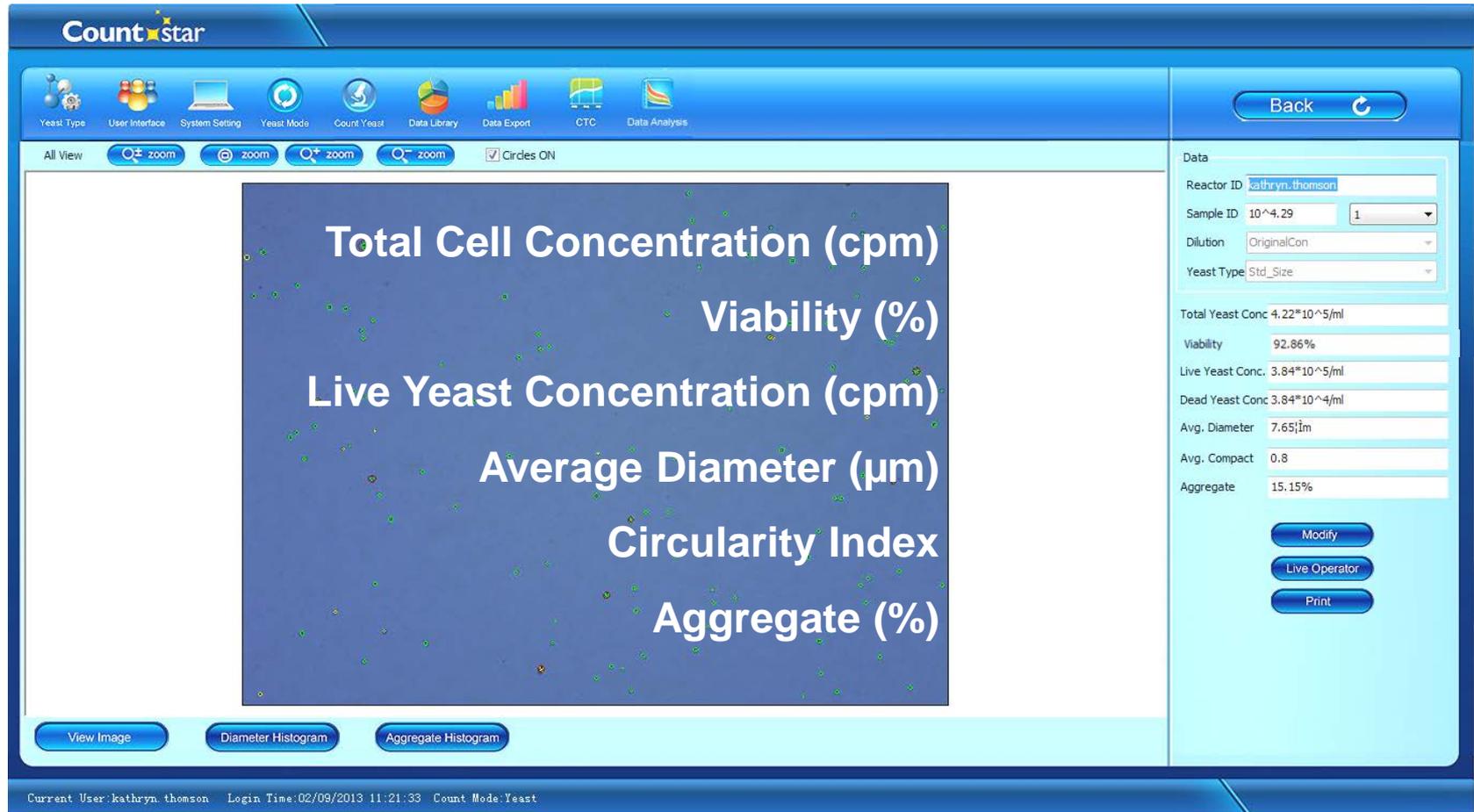


Aber Countstar

- ▶ Automated cell counter
- ▶ Methylene blue or MV
- ▶ Five chamber disposable slides
- ▶ Additional information on yeast



Information provided by the Countstar



The screenshot displays the Countstar software interface. The main window shows a microscopic image of yeast cells with overlaid text labels for various parameters: Total Cell Concentration (cpm), Viability (%), Live Yeast Concentration (cpm), Average Diameter (μm), Circularity Index, and Aggregate (%). The interface includes a top navigation bar with icons for Yeast Type, User Interface, System Setting, Yeast Mode, Count Yeast, Data Library, Data Export, CTC, and Data Analysis. A right-hand panel contains a 'Data' section with input fields for Reactor ID (kathryn.thomson), Sample ID (10^4.29), Dilution (OriginalCon), and Yeast Type (Std_Size). Below these are numerical results for Total Yeast Conc (4.22*10^5/ml), Viability (92.86%), Live Yeast Conc (3.84*10^5/ml), Dead Yeast Conc (3.84*10^4/ml), Avg. Diameter (7.65 μm), Avg. Compact (0.8), and Aggregate (15.15%). Buttons for Modify, Live Operator, and Print are located at the bottom of the data panel. The bottom status bar shows 'Current User: kathryn.thomson Login Time: 02/09/2013 11:21:33 Count Mode: Yeast'.

Countstar

Yeast Type User Interface System Setting Yeast Mode Count Yeast Data Library Data Export CTC Data Analysis

All View zoom zoom zoom zoom Circles ON

Total Cell Concentration (cpm)
Viability (%)
Live Yeast Concentration (cpm)
Average Diameter (μm)
Circularity Index
Aggregate (%)

View Image Diameter Histogram Aggregate Histogram

Data

Reactor ID:

Sample ID:

Dilution:

Yeast Type:

Total Yeast Conc: 4.22*10^5/ml

Viability: 92.86%

Live Yeast Conc: 3.84*10^5/ml

Dead Yeast Conc: 3.84*10^4/ml

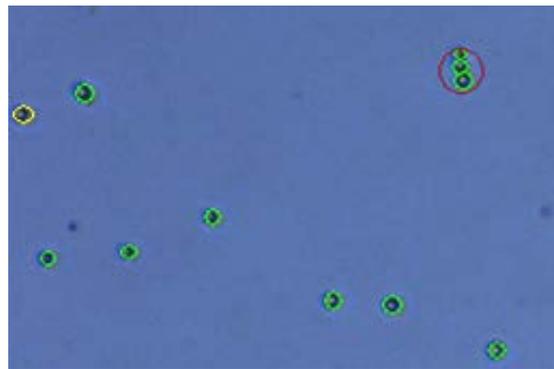
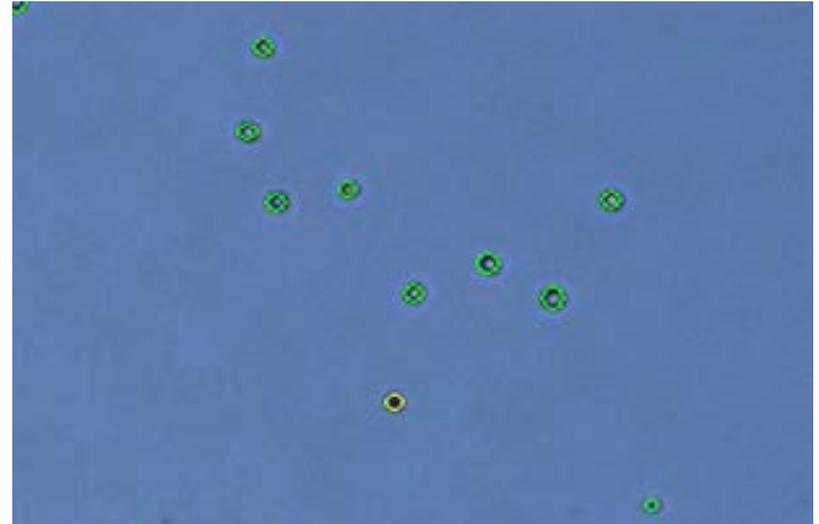
Avg. Diameter: 7.65 μm

Avg. Compact: 0.8

Aggregate: 15.15%

Current User: kathryn.thomson Login Time: 02/09/2013 11:21:33 Count Mode: Yeast

Dead vs Alive cells



Example with propagating yeast

Countstar

Yeast Type User Interface System Setting Yeast Mode Count Yeast Data Library Data Export CTC Data Analysis

All View zoom zoom zoom zoom Circles ON

Back

Data

Reactor ID john

Sample ID YPR2 1

Dilution 1:1

Yeast Type Std_Size

Total Yeast Conc 1.78×10^7 /ml

Viability 97.31%

Live Yeast Conc. 1.73×10^7 /ml

Dead Yeast Conc 4.8×10^5 /ml

Avg. Diameter 7.28µm

Avg. Compact 0.72

Aggregate 86.48%

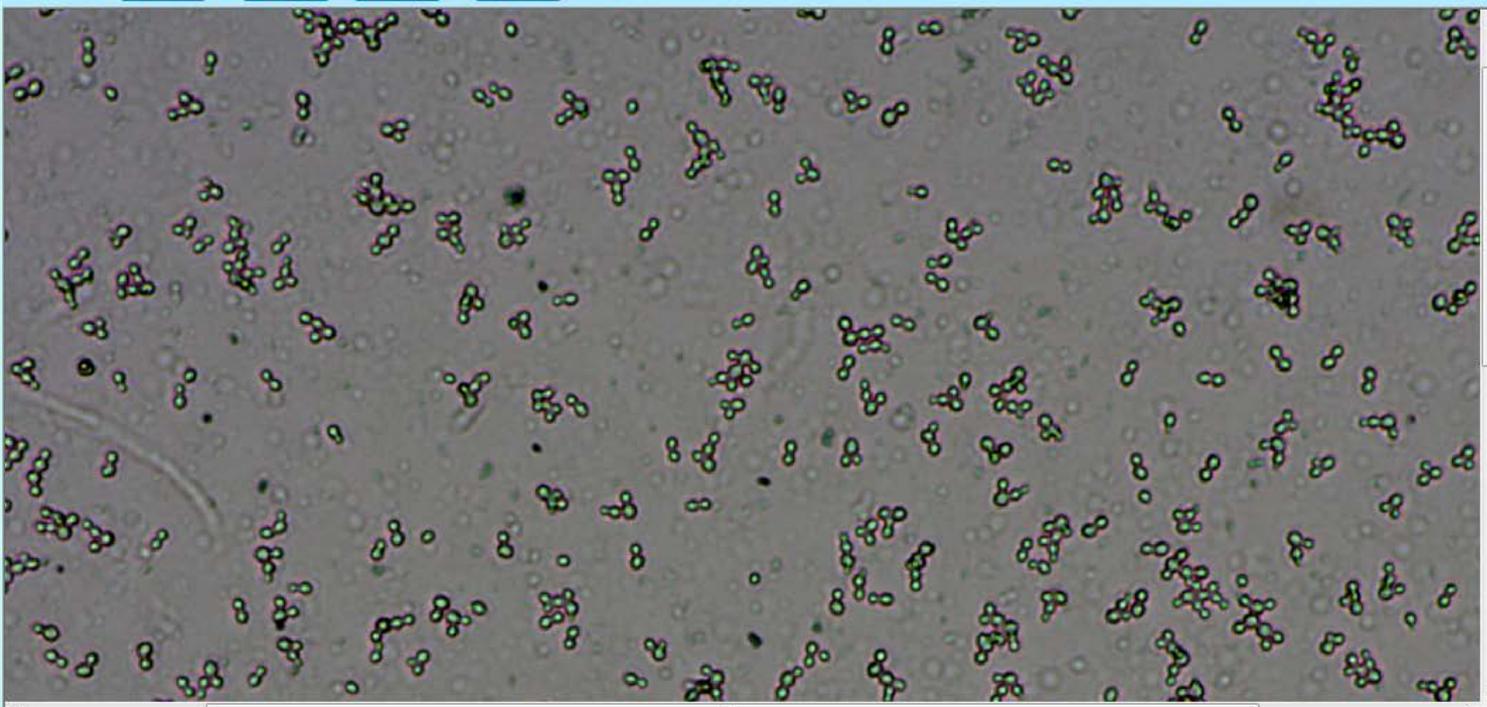
Modify

Live Operator

Print

View Image Diameter Histogram Aggregate Histogram

Current User: john Login Time:01/03/2015 21:32:19 Count Mode:Yeast



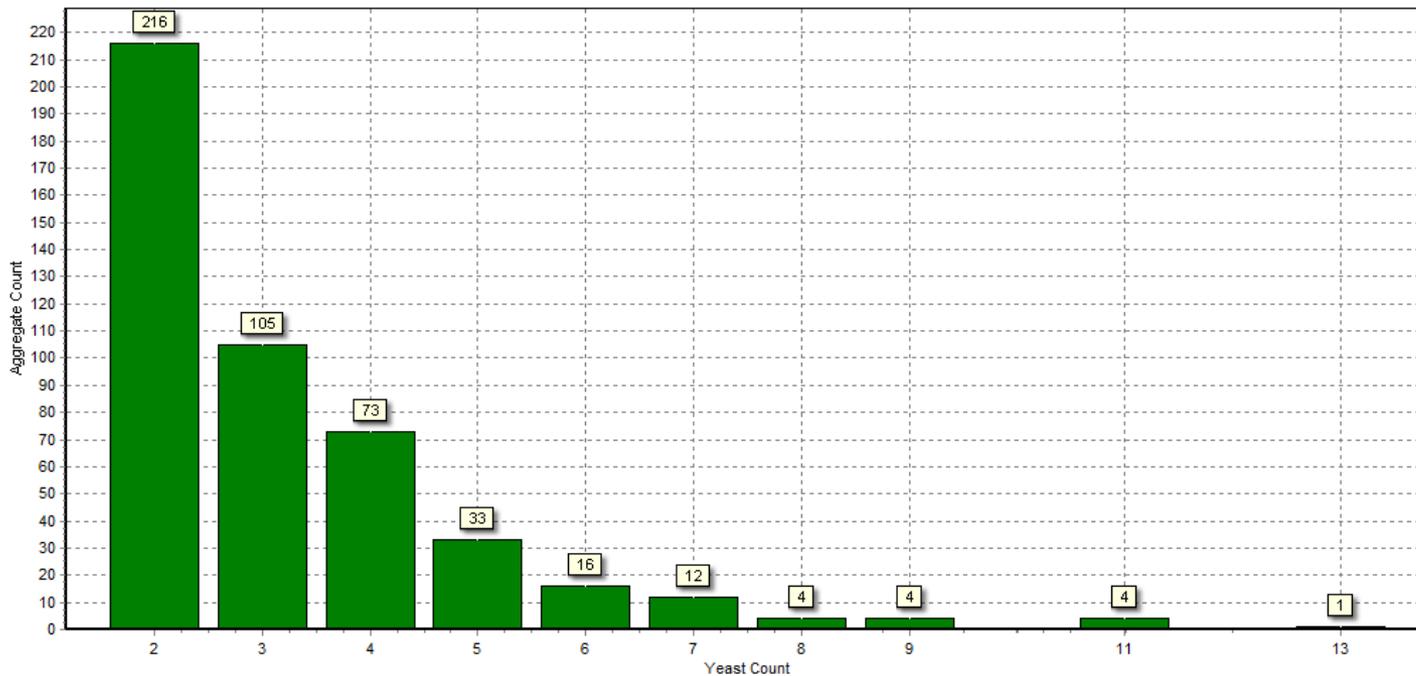
Data
Reactor ID john
Sample ID YPR2 1
Dilution 1:1
Yeast Type Std_Size

Total Yeast Conc	1.78*10 ⁷ /ml
Viability	97.31%
Live Yeast Conc.	1.73*10 ⁷ /ml
Dead Yeast Conc	4.8*10 ⁵ /ml
Avg. Diameter	7.28µm
Avg. Compact	0.72
Aggregate	86.48%



Back

Aggregate Histogram



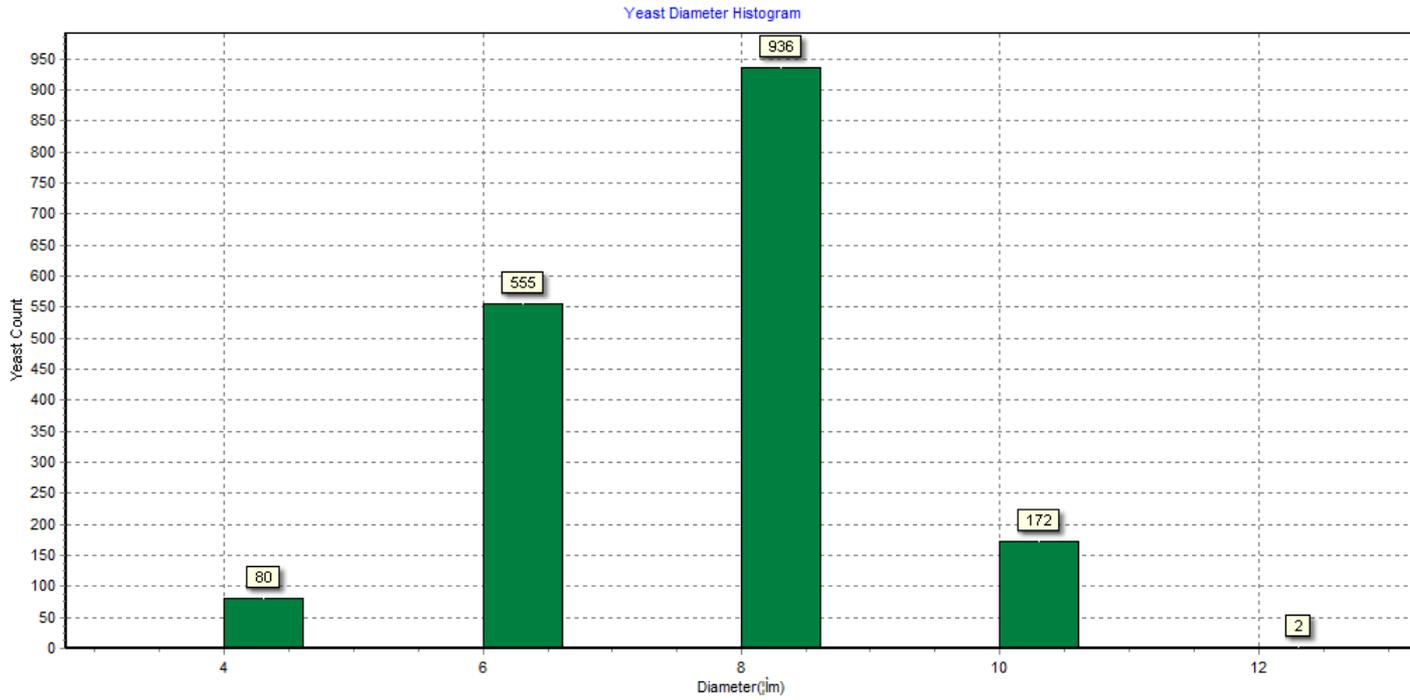
Data

Reactor ID: john
 Sample ID: YPR2 1
 Dilution: 1:1
 Yeast Type: Std_Size

Total Yeast Conc: 1.78×10^7 /ml
 Viability: 97.31%
 Live Yeast Conc: 1.73×10^7 /ml
 Dead Yeast Conc: 4.8×10^5 /ml
 Avg. Diameter: 7.28 μm
 Avg. Compact: 0.72
 Aggregate: 86.48%

Modify
 Live Operator
 Print

View Image Diameter Histogram Aggregate Histogram



Data

Reactor ID:

Sample ID: YPR2

Dilution: 1:1

Yeast Type: Std_Size

Total Yeast Conc	1.78*10 ⁷ /ml
Viability	97.31%
Live Yeast Conc.	1.73*10 ⁷ /ml
Dead Yeast Conc	4.8*10 ⁵ /ml
Avg. Diameter	7.28μm
Avg. Compact	0.72
Aggregate	86.48%

- [Modify](#)
- [Live Operator](#)
- [Print](#)

Objectives of work

- ▶ To see if the gold standard stain can be retained, while automating the process.
- ▶ Investigate whether consistency, labour time and inter-operator effects were improved.
- ▶ Reveal how well an automated machine that uses methylene blue can be applied in the brewing industry.

Methods I

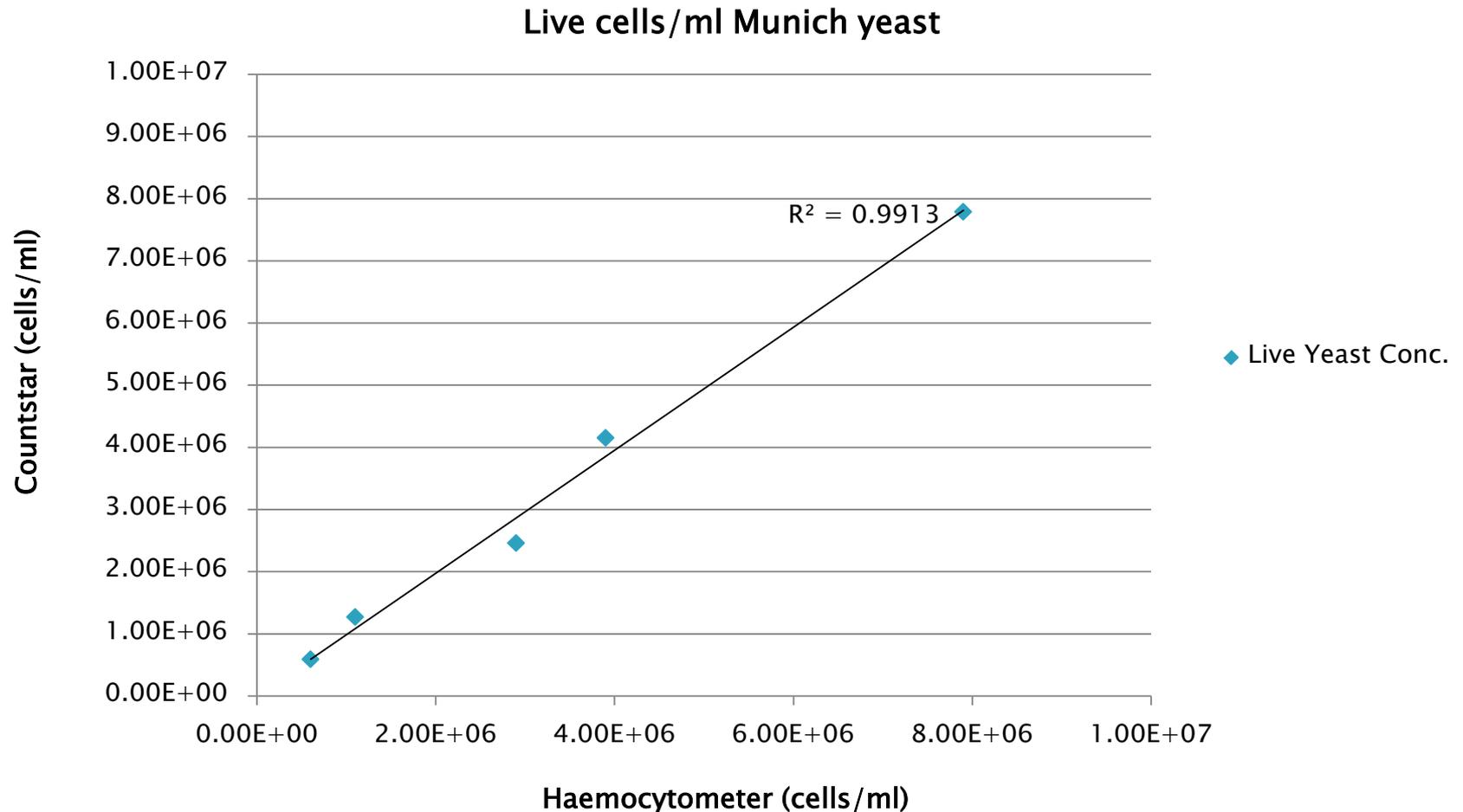
- ▶ Dried brewer's Yeast
 - In house testing with a range of dried brewer's yeast.
 - Samples analysed with the Aber Countstar and haemocytometer.
 - Live cell concentration (cells/ml) recorded using each method
 - Results directly compared.

Results I – Dried brewer's Yeast

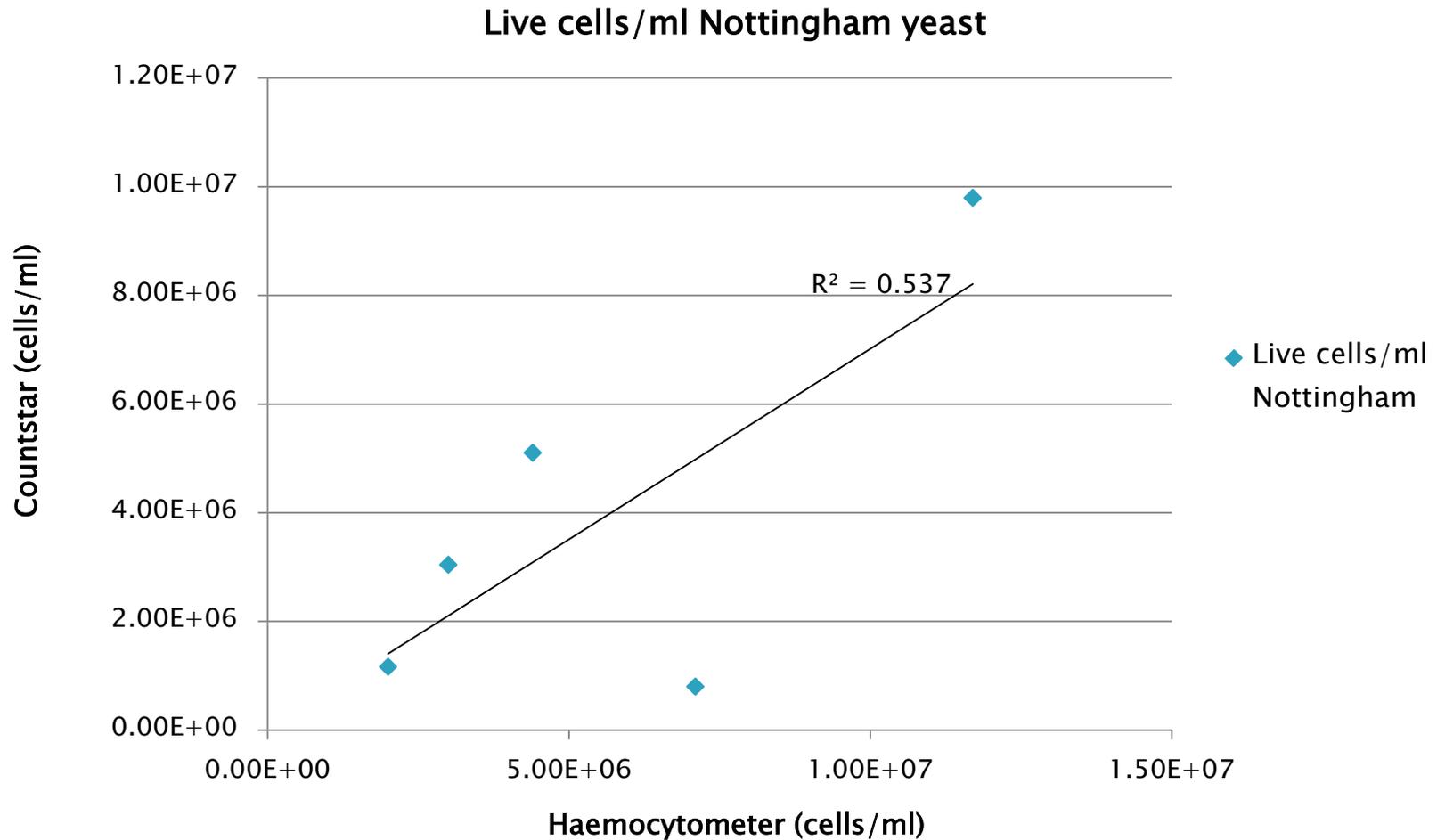
Saccharomyces cerevisiae

Yeast strain name	Type	R ² value
Nottingham	Ale yeast	0.537
BRY-97 American West Coast	Ale yeast	0.957
Belle Saison	Belgian Ale yeast	0.985
EC-118	Champagne yeast	0.919
Munich	Wheat beer yeast	0.991
Kolsch A	German Beer yeast	0.576

Results I – Dried brewer's Yeast.

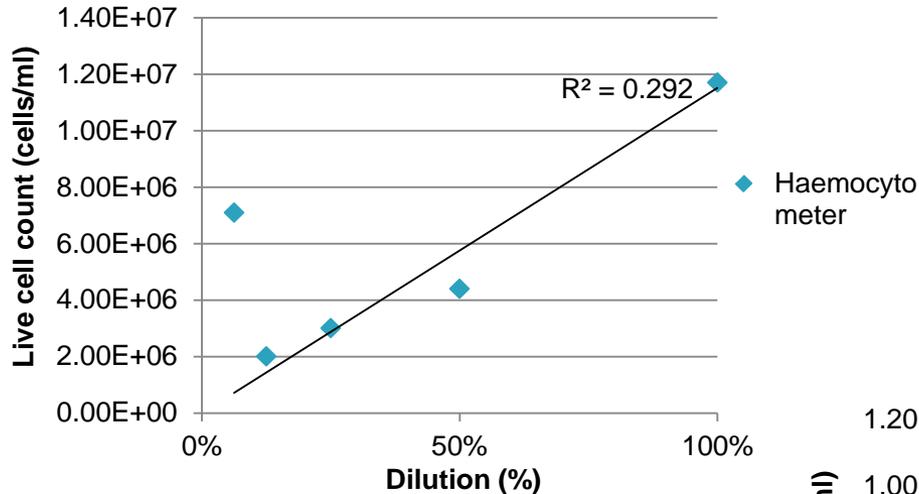


Results I – Dried brewer's Yeast

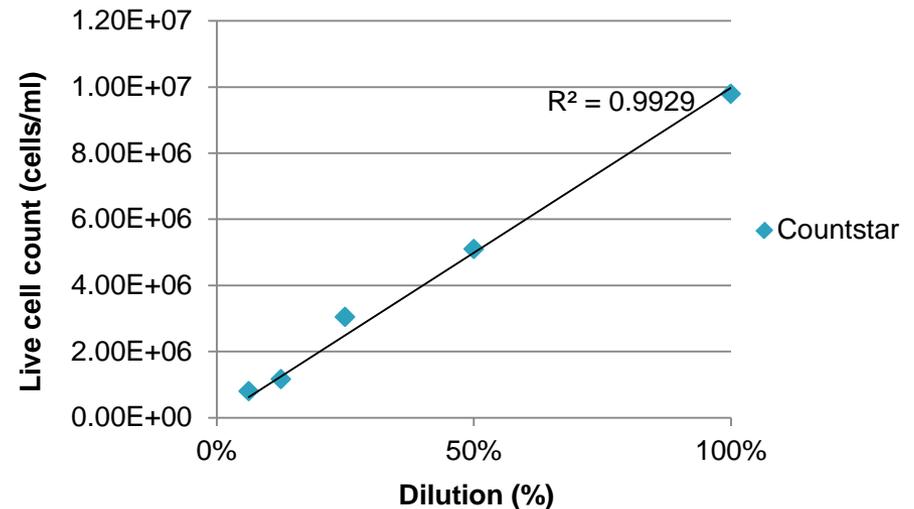


Results I – Dried brewer's Yeast

Haemocytometer Live cell count Nottingham yeast



Countstar Live cells/ml Nottingham yeast

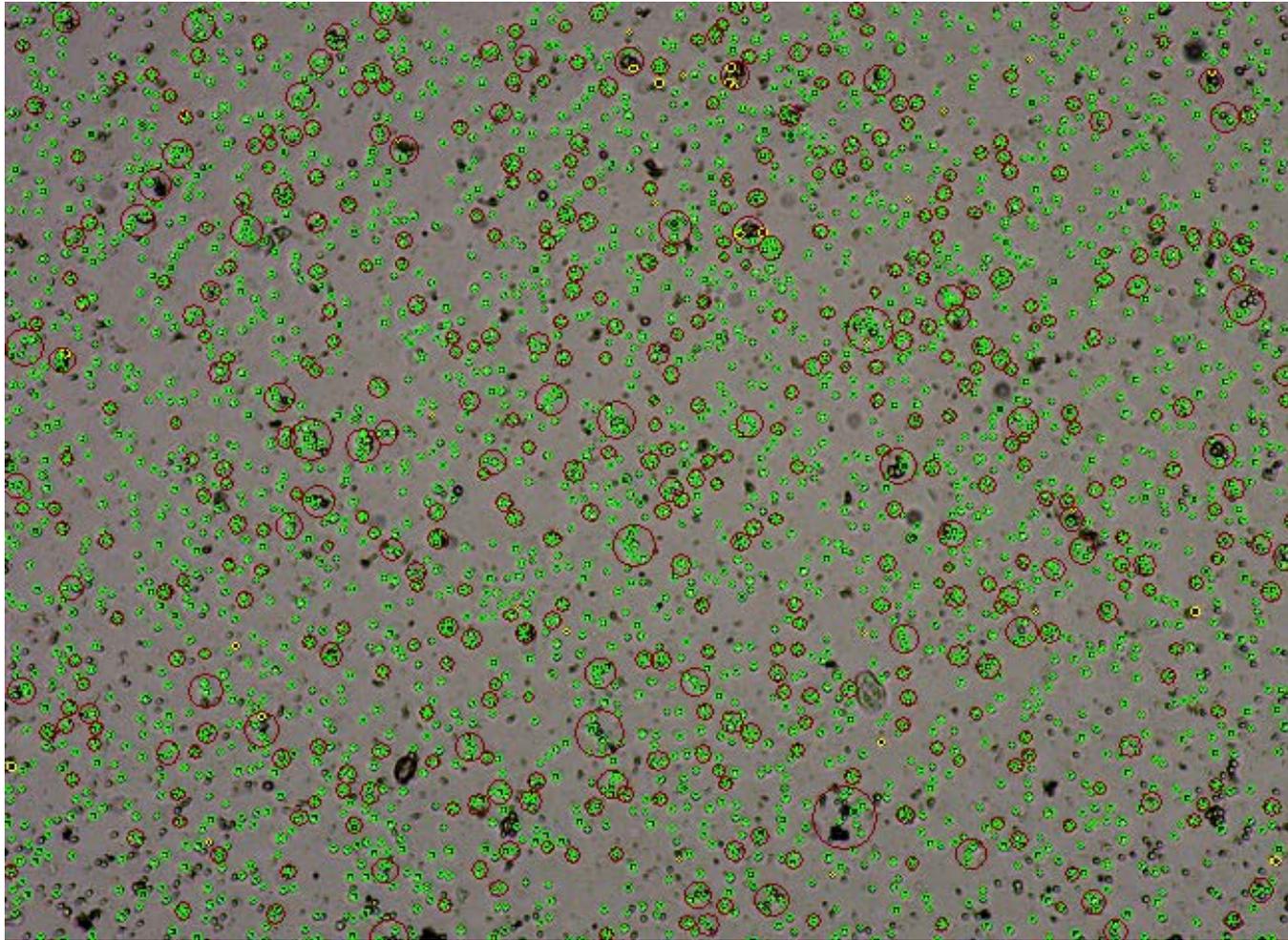


Methods II

- ▶ Wheat beer
 - Bottle conditioned
 - Analysed using haemocytometer and Countstar
 - Range of concentrations tested with 2-fold serial dilutions.
 - Live cell count using both methods assessed and compared.



Results II – Bottled Wheat Beer

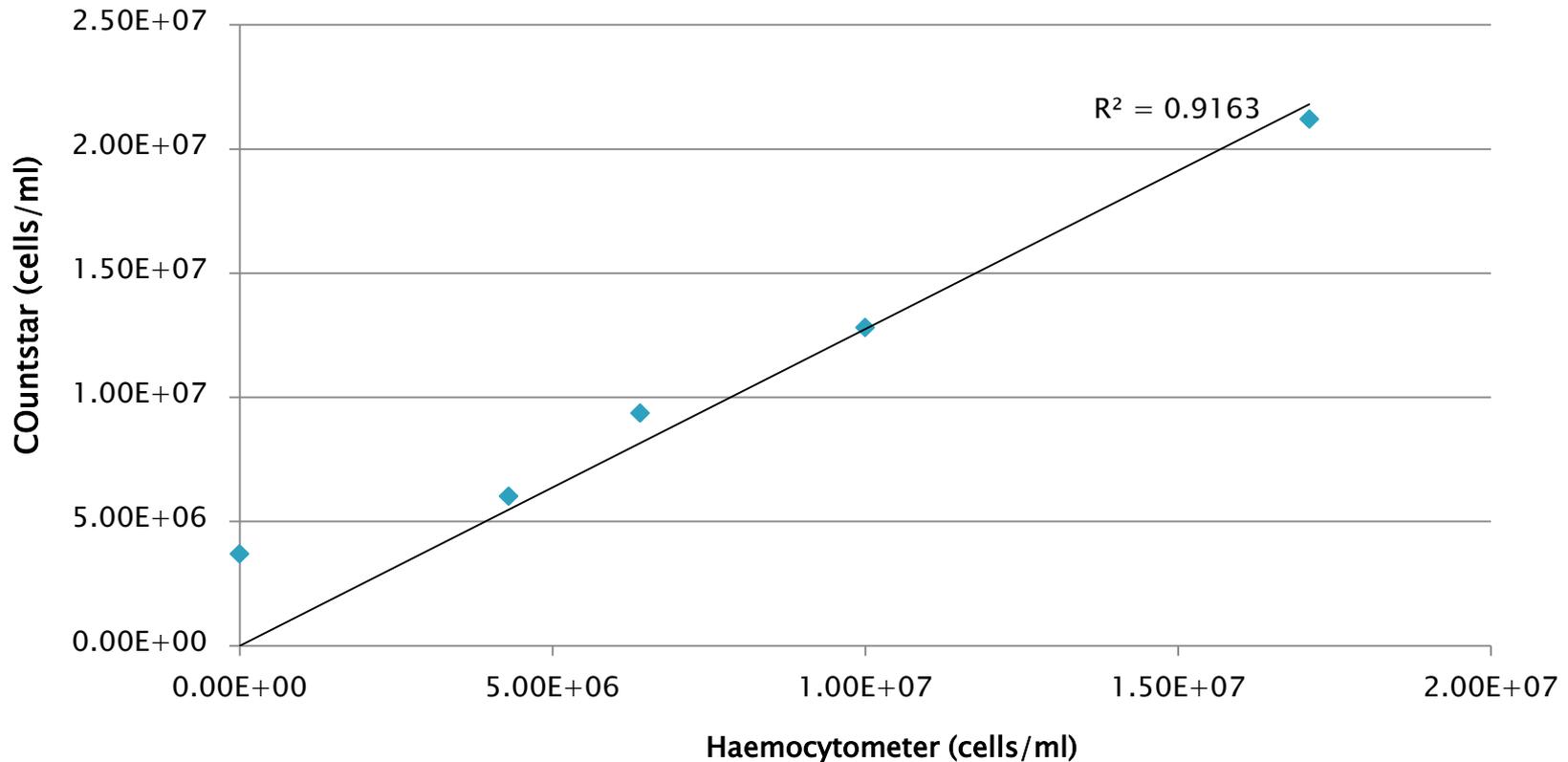


Results II – Bottled Wheat Beer



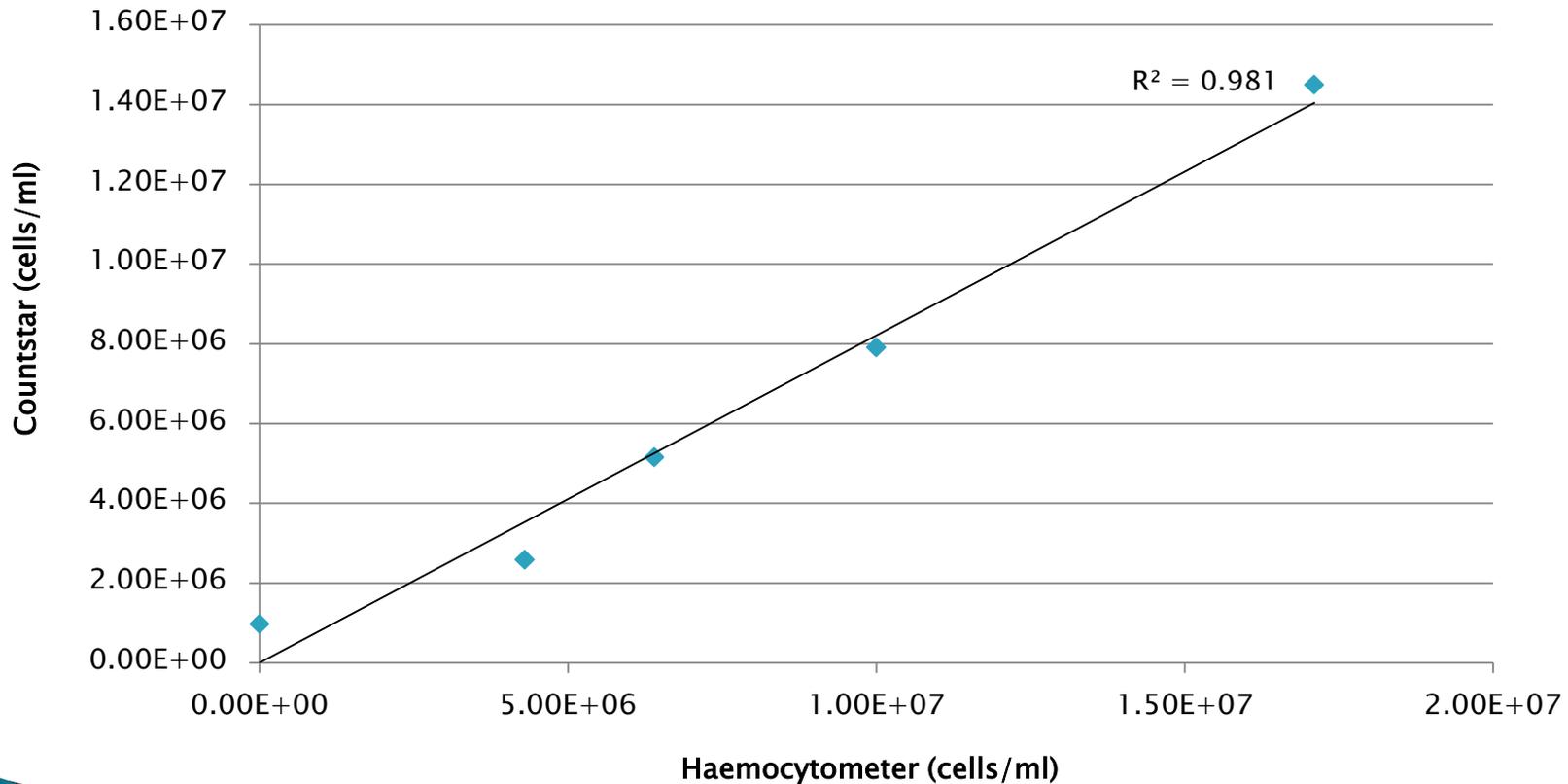
Results II - Bottled Wheat Beer

Haemocytometer vs Default Parameters Countstar



Results II – Bottled Wheat Beer

Haemocytometer vs Countstar with Altered Parameter settings live cells/ml



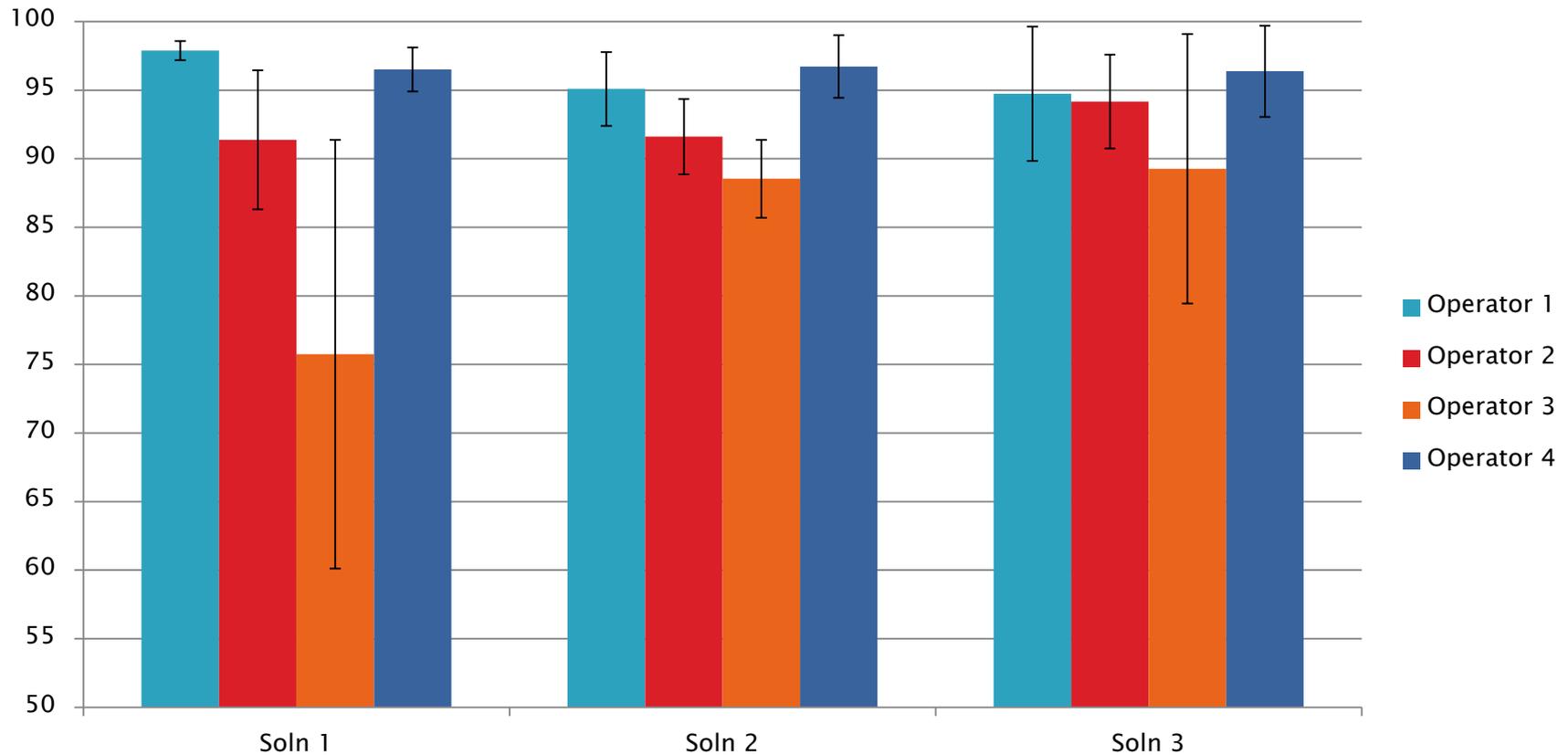
Methods III

- ▶ Inter-Operator Variability
 - Baker's yeast.
 - Three separate solutions.
 - Four operator's (different abilities) measured cell count and viability for each solution three times.
 - An average for each operator was taken for each of the three solutions.



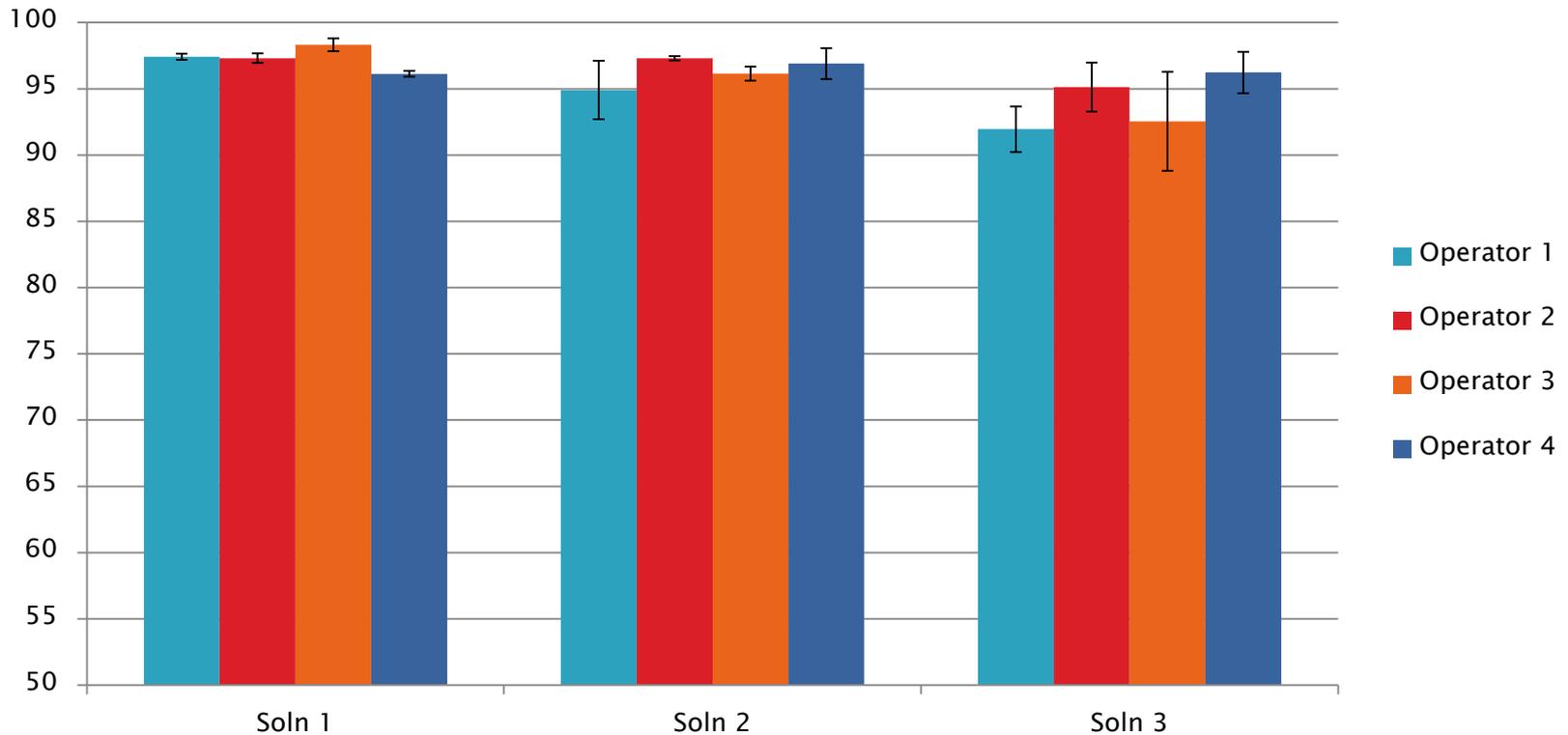
Results III – Inter-operator Variability

Haemocytometer % viability



Results III -Inter-operator Variability

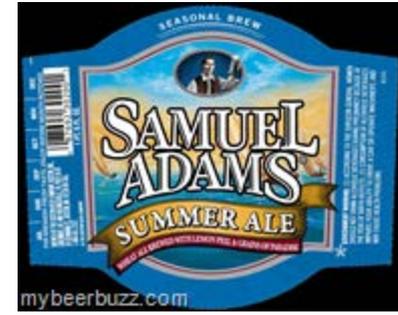
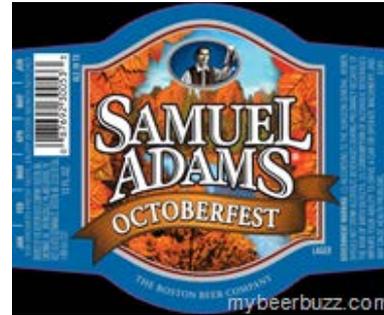
Counstar % viability



Methods IV

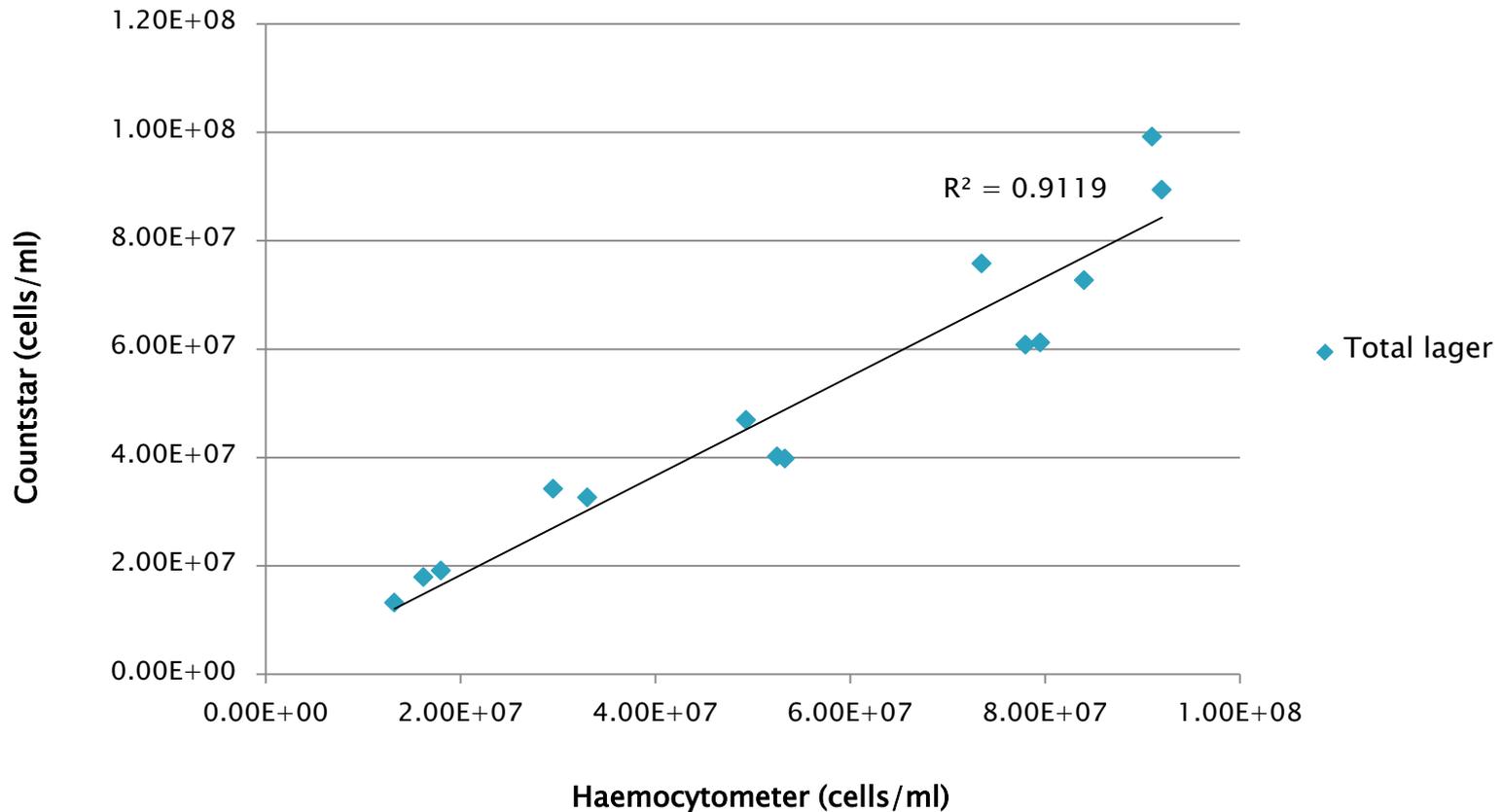
▶ Boston Beer

- Propagation yeast
 - Total cell count most important in propagation yeast using Countstar and manually.
 - 11 samples of ale and 11 of lager yeast were analysed.
- Harvest yeast
 - Viability% recorded.
 - 27 samples of ale and 20 samples of lager yeast.
- Fermenter samples
 - Total cell counts recorded manual counts and Countstar.
 - 3 samples of ale and 3 of lager yeast were analysed.



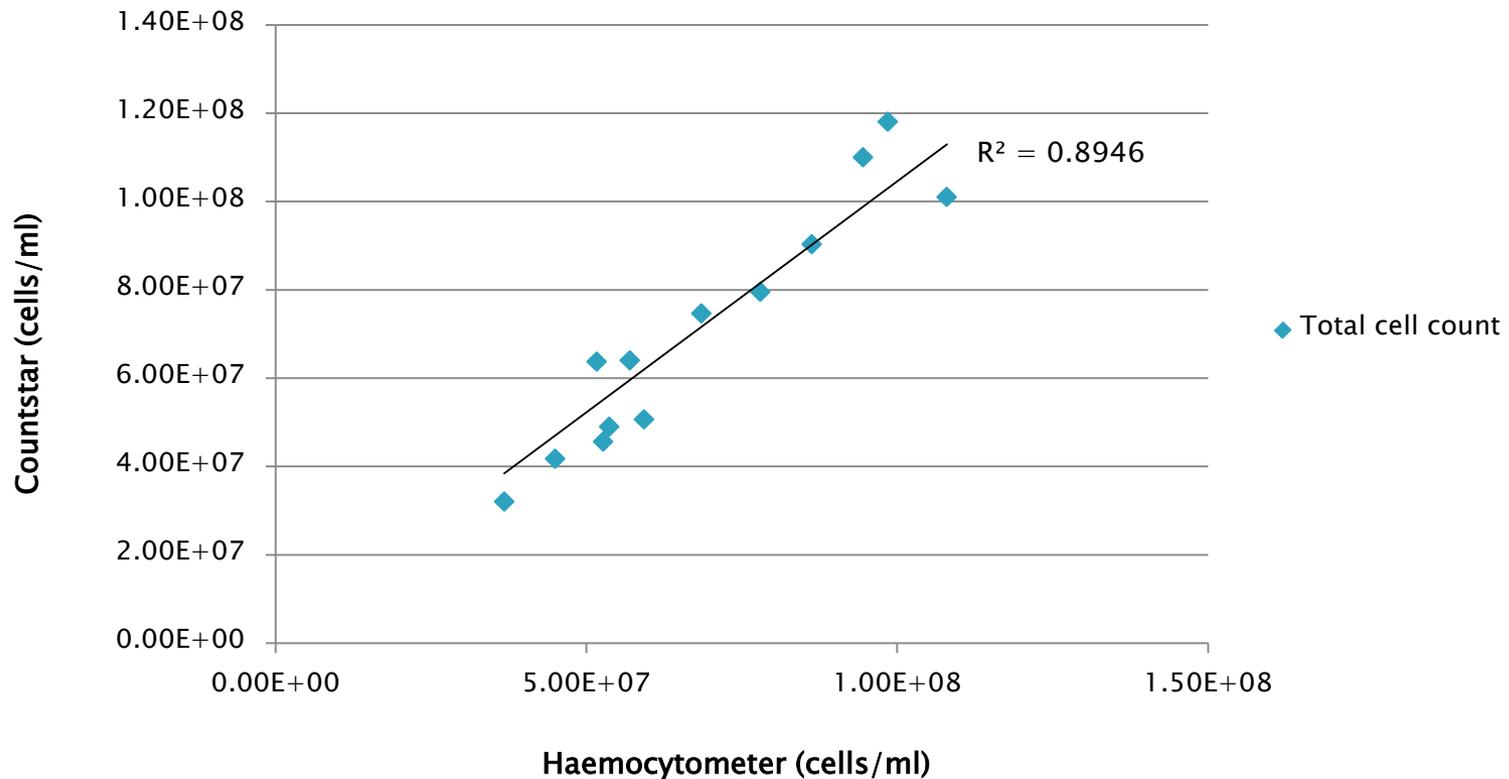
Result IV – Boston Beer

Total lager cell count recorded with Countstar and Haemocytometer



Results IV – Boston Beer

Total cell count of an ale yeast as recorded with Countstar and Haemocytometer



Conclusions

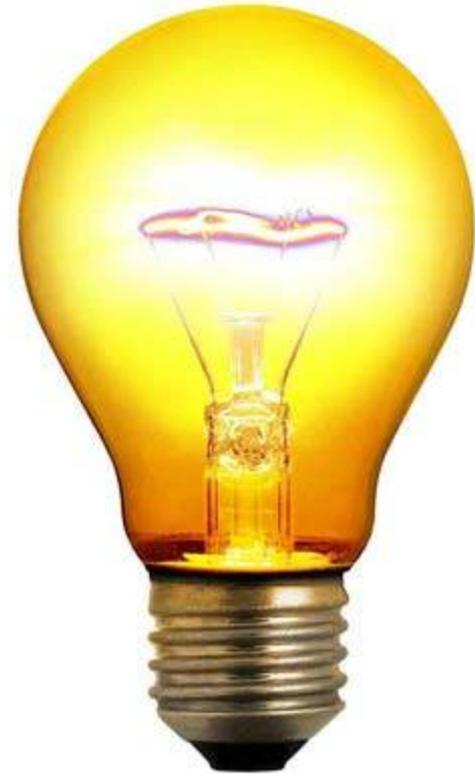
- ▶ The Aber Countstar can accurately perform cell counts with a range of dried brewing yeasts.
- ▶ Shown to be more reliable with some of these yeasts than haemocytometer readings.
- ▶ The Countstar is able to discount trub (Wheat beer).
- ▶ Lower deviation between repeats for one operator.
- ▶ Lower inter-operation discrepancies displayed when multiple operators analysed samples with the Countstar than when performed manually.
- ▶ Able to distinguish buds in propagation yeast.

Possible Impact to Brewing

- ▶ Save manual labour time
- ▶ Allows multiple operators to analyse samples with much lower deviation.
- ▶ Allows different multiple breweries within group to have “ the same microbiologist’s interpretation of count and viability”
- ▶ Images of yeast can be compared (across different site too) to reference images for possible cross contamination or changes in yeast morphology

Ongoing Research.

- ▶ Budding cells
- ▶ Use of technology in wine and spirits production.



Acknowledgements

- ▶ Katy Thomson, Aber Instruments
- ▶ Dr Urs Wellhoener, Boston Beer Company



Thank you for Listening.

Any Questions?